REVIEW

CYP2C19 and CYP2D6 genotypes in Pacific peoples

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The study of pharmacogenetic variants in populations which reside in Oceania has been focused mainly on CYP2C19 and CYP2D6. Statements about the high prevalence of CYP2C19 no function genotype in 'Pacific Islanders' can be found in the literature. This review article summarizes the published information about these pharmacogenes in this geographical region and highlights the differences observed between Melanesian and Polynesian populations. It is not appropriate to combine the prevalence data of pharmacogenetic variants, particularly CYP2C19, across this region. Indeed, apocryphal assumptions about CYP2C19 no function alleles and possible effect on the therapeutic activity of clopidogrel are unhelpful and reiterate the importance of assessing the individual patient rather than relying on inappropriate ethnicity-based assumptions for drug dosing decisions.

Introduction

The Western Pacific region broadly encompasses countries of Southeast Asia, such as the east coast of mainland China, Korea, Taiwan and Japan as well as the islands of Oceania. This latter region includes the large islands of Papua New Guinea, Australia and New Zealand as well as a myriad of smaller Pacific islands. There are more than 25 000 Pacific islands but many of these are uninhabited. The Pacific islands are divided into the regions of Micronesia (including Palau, Marianas, Caroline and Marshall Islands and Kiribati); Melanesia, an area extending from Papua New Guinea, via an archipelago of islands to New Caledonia and Fiji; and a vast area called Polynesia which extends from Hawai'i, Rapanui to the most southerly island in the Polynesian group, Aotearoa (New Zealand).

What is the difference between the Pacific peoples of Melanesia and Polynesia?

There is evidence that there have been multiple waves of migration of mankind into the Western Pacific region.

Archaeological evidence suggests that humans crossed land bridges and arrived in Melanesia more than 45 000 years ago. Whole genome sequencing also suggests that anatomically modern humans may have coexisted and possibly admixed with Denisovan-like archaic hominids in this region [1]. At the end of the last ice age (<10000 years ago) the populations of Papua New Guinea, Melanesia and Aboriginal Australians became geographically isolated from the later wave of migration of anatomically modern man into Southeast Asia and Indonesia. This links with the genomic ancestry cline observed between Melanesia and island Southeast Asia [2], which broadly corresponds to the biogeographic faunal discontinuity known as the Wallace line. A much later (~3000 years ago) wave of migration by sea, possibly from a region close to modern day Taiwan, is thought to have led to colonization of some of island Melanesia as well as the further expansion into the remote Polynesian islands such as Hawai'i, Rapanui and Aotearoa (New Zealand). Oral genealogical information indicates that the arrival of Maori in New Zealand occurred about 1000-800 years ago and more than forty separate waka (double hulled canoes) undertook this long journey. Many of the iwi (tribes) of Maori can trace their origins to the captains and crew of these waka.



Much of this human history of the Pacific can be ascertained from archaeological and linguistic studies. However, use of genome-wide single nucleotide polymorphism (SNP) data have also inferred these demographic relationships [3]. Hence, there are not only geographical but also substantial cultural and ethnic differences across this region, particularly between the populations residing in Melanesia and Polynesia. The complexities of ancestral demographics and geographic isolation, admixture with European settlers, as well as the possible confounding effects of natural disasters, especially on the small volcanic islands and atolls of the Pacific, can result in substantial bottleneck effects and may underpin the differences in the prevalence of genetic polymorphisms in drug-metabolizing enzymes between individual populations across Oceania.

CYP2C19

Oceania is considered to have the highest frequency of no function *CYP2C19* individuals of any geographical region. Consequently up to 58% of individuals from this region are expected to have a no function CYP2C19 'poor metabolizer' phenotype [4]. However, it is important to note that these values are determined from ethnically diverse populations from geographically distant regions

(Table 1). These high prevalence values are particularly influenced by the data reported from more than 6978 individuals located in Melanesia, i.e. Papua New Guinea and Vanuatu [5–10] and also Australian aboriginal people [11]. Based on the data from Vanuatu, it was predicted 'that the poor metabolizer genotype is common throughout Polynesia and Micronesia' [7]. However, CYP2C19 status has only been assessed in 246 Polynesian individuals (3.4% of all Oceanian subjects tested) and there are no reports in Micronesian individuals. Moreover, many of these reports in Polynesian subjects have limitations, such as assessment of CYP2C19 in cohorts of patients, e.g. Lupus nephritis and cardiovascular disease [12, 13], or have only assessed phenotype [14, 15]. Even with these limitations, the average prevalence of inherited CYP2C19 poor metabolizer status in Polynesia is 8% and is substantially lower than the mean reported across Melanesia (50.1%; Table 1). In addition, the CYP2C19*3 allele appears to be relatively uncommon in Polynesians compared with Melanesians, with an allele frequency of 0.04 vs. 0.19 (Table 1). The CYP2C19*4 no function allele has not been assessed in Pacific peoples. The prevalence of the increased function allele CYP2C19*17 is unclear (range 0.0-0.11) as two publications have assessed this variant and both these are from small subgroup observations in patient cohorts [12, 13].

Table 1Prevalence of CYP2C19 no function alleles reported in different populations and geographical regions across Oceania

Geographical region/Ethnicity	CYP2C19*2	CYP2C19*3	Homozygous no function CYP2C19 (poor metaboliser) ^a	Number of subjects	Reference
Papua New Guinea (>6 individual locations)	0.45	0.16	36%	401	[9]
	0.37	0.34	49%	47 ^b	[5]
	0.45	0.19	40%	172	[10]
Vanuatu (>24 individual locations)	0.71	0.13	70.8%	493	[6]
	0.63	0.14	61%	5538	[7]
	0.57	0.25	68%	100	[8]
Aboriginal Australians (1 location)	0.36	0.14	25.6%	227	[11]
Summary ^c of Melanesia	0.51	0.19	50.1%	6978	
Samoan, Tongan, Cook Islander, Niuean ^d	n.d.	n.d.	13% ^e	59	[15]
Samoan, Tongan, Fijian, Cook Islander ^{d,f}	0.14	0.07	4%	14	[12]
Maori and 'Pacific islanders'dg	0.29	n.d.	8%	70	[13]
Maori	n.d.	n.d.	7% ^d	43	[14]
Maori	0.24	0.017	8%	60	[22]
Summary ^c of Polynesia	0.22	0.04	8%	246	

n.d., not determined

^aIncludes compound heterozygotes (*2/*3)

^bTwo locations excluded due to very small sample size

^cSummary are mean values

^dSelf-identified ethnicity, resident in NZ

^ePhenotyped only, no genotype information collected

fLupus nephritis

^gCardiovascular disease



CYP2C19 and clinical outcomes in Pacific peoples

CYP2C19 poor metabolizers may have a poor therapeutic response to the chemoprophylactic antimalarial prodrug proguanil. Asymptomatic malaria-infected Vanuatuan children (Melanesia), who were CYP2C19 homozygous for no function alleles, had a significantly lower formation of cycloguanil, the active metabolite of proguanil, than individuals who were not carriers of no function alleles. Indeed many of these poor metabolizer individuals had no detectable cycloguanil formation [8]. However, the response of Plasmodium vivax or Plasmodium falciparum infection to treatment with proguanil did not correlate with no CYP2C19 genotype [16]. It is possible that adequate blood concentrations of proguanil were achieved in these CYP2C19 no function children who had mild or asymptomatic infection as the parent compound has some intrinsic (albeit weak) antimalarial activity. Thus, the high prevalence of this genetic polymorphism in this geographic location may have no clinical consequence in this particular context.

The presence of no function CYP2C19*2 is strongly associated with an increased risk of a stent thrombosis when on clopidogrel therapy [17]. Maori and Pacific Island patients do appear to have a significantly higher rate of 'high on clopidogrel platelet reactivity' compared to New Zealand Europeans (57% vs. 35.9%, [18]). However, this study did not assess CYP2C19 genotype. In a separate study [19], although not directly measured, a higher prevalence of CYP2C19*2 allele frequency was assumed in Maori (0.24) and Pacific islanders (0.45) compared to European New Zealanders (0.15). Based on these prevalence values, no CYP2C19*2 gene dose association was observed for rates of stent thrombosis across these ethnicities (event rates: 0.3%, 0.3% and 0.2% for European, Maori and Pacific Islanders, respectively), or for other events, such as bleeding, myocardial infarction, stoke or cardiovascular death. Direct assessment of CYP2C19 genotype was undertaken in a more recent study of 312 New Zealand patients, of whom 70 were Maori and Pacific islanders [13]. The CYP2C19*2 allele could only explain 3-4% of the variability of on-treatment platelet reactivity and this study reiterated the known importance of clinical variables such as comorbidities and co-medications in clopidogrel outcomes, which accounted for 18% of this variability. There is a high rate of type 2 diabetes in Maori and Pacific Islanders in New Zealand with rates reported to be 218-370 per 100 000 compared to 97 per 100 000 in the total New Zealand population (http://www.stats.govt.nz/). Poor cardiovascular outcomes are a known consequence of chronic diabetes and Maori and Pacific islanders have a higher rate of myocardial infarction and cardiovascular death on clopidogrel treatment than European patients [19]. Importantly, regardless of ethnicity, only diabetes and CYP2C19*2 were independent contributors to poor control of platelet reactivity on clopidogrel treatment in this New Zealand cohort [13].

In 2014 the Attorney General of the state of Hawai'i raised concerns that certain pharmaceutical suppliers of clopidogrel had failed 'to disclose that a significant proportion of patients in Hawai'i had genetic polymorphism in the [CYP2C19 gene]' [20]. It is important to note that the assumption that the

prevalence of CYP2C19 poor metabolism is high in Polynesians is not supported by the current literature, and the direct assessment of this genotype in Hawai'ian Polynesians has not been reported. Despite this lack of evidence, articles raise the concern that 'the standard 75 mg dose of clopidogrel is not efficacious for Hawaiians' [21]. These authors also state 'As a total population some 150,000-300,000 Hawaiians will have a reduced enzymatic capacity to convert clopidogrel to its active metabolite'. Such apocryphal statements are clearly unhelpful, particularly as data from a New Zealand cohort of Polynesians support the known importance of clinical comorbidities in clopidogrel treatment outcomes for patients with acute coronary syndromes.

CYP2D6 in Pacific peoples

The CYP2D6 poor metabolizer phenotype has been reported at 5% frequency in Maori, using debrisoquine as a probe drug [14]. CYP2D6 allelic variants that either lack enzyme activity (*4,*5) or result in decreased activity (*41) appear to be present at relatively low frequency in Maori compared to Caucasian Europeans [22]. An absence of the CYP2D6 poor metabolizer phenotype has been reported in 'South Pacific Polynesians' [15] and a low frequency of CYP2D6 poor metabolizer status (1%) is also predicted from the assessment of CYP2D6*4, *10 and *41 alleles in subjects from Papua New Guinea [10] and is based on genotype data. The poor metabolizer phenotype is also predicted to be relatively low in (0.4%) in Australian Aboriginal people [11]. Furthermore, CYP2D6 genotype data of a small cohort of patients in Auckland suggests that the prevalence of no/decreased function CYP2D6 alleles may be lower in Maori and Pacific islanders than in New Zealand Europeans [23]. These limited data suggest that CYP2D6 no function genotype is not common in Polynesian or Melanesian individuals (as reviewed in [24]). However, a proportion of individuals in some of these studies [10, 23] could not be assigned a genotype (no call) suggesting additional unidentified sequence variations or novel haplotype(s) of known SNPs within these populations. CYP2D6 gene duplication (CYP2D6*2xN), associated with an ultrarapid metabolizer phenotype, was observed at 0.12 frequency in subjects from Papua New Guinea [10]. This duplication, however, was absent in Australian aborigines [11] and little is known about the prevalence of this gene duplication in Polynesians and other Pacific Islanders. There do not appear to have been any clinical studies to assess different clinical outcomes in Pacific peoples based on CYP2D6 genotype.

The prevalence of some other CYP enzymes, such as CYP2C9 and CYP2A6 have been reported in Maori [22], CYP2E1 in Aboriginal Australians [11] and CYP2B6 in Papua New Guinea [25]; however, it is not possible to compare these prevalence rates across the geographic locations and ethnicities of the Pacific.

Conclusions

The prevalence of CYP2D6 no function phenotype is likely to be low in populations of Polynesian and Melanesian ancestry,



although this is based on limited evidence. In contrast, substantial data indicate that CYP2C19 no function variants are very common in Melanesians, but these variants appear to be much less frequent in the small number of Polynesian individuals tested. Based on the demographics of human history in the Pacific region, it is clear that substantial differences exist between people of Melanesian and Polynesian ancestry. It is not appropriate to combine the prevalence data of pharmacogenetic variants, particularly CYP2C19, across this vast region. Statements about the high prevalence of CYP2C19 no function genotype in 'Pacific Islanders' are apocryphal and unhelpful. Assumptions about the presence of no activity pharmacogenetic variants based on apparent and/or self-reported ethnicity is never appropriate. Dosing decisions, when justified for the safe and effective use of a particular medication, should be based on the known genotype of the individual patient.

Competing Interests

There are no competing interests to declare.

References

- 1 Veeramah KR, Hammer MF. The impact of whole-genome sequencing on the reconstruction of human population history. Nat Rev Genet 2014; 15: 149–62.
- 2 Cox MP, Karafet TM, Lansing JS, Sudoyo H, Hammer MF. Autosomal and X-linked single nucleotide polymorphisms reveal a steep Asian–Melanesian ancestry cline in eastern Indonesia and a sex bias in admixture rates. Proc Roy Soc Lond B Biol Sci 2010; 277: 1589–96.
- **3** Wollstein A, Lao O, Becker C, Brauer S, Trent RJ, Nürnberg P, *et al.* Demographic history of Oceania inferred from genome-wide data. Curr Biol 2010; 20: 1983–92.
- **4** Fricke-Galindo I, Céspedes-Garro C, Rodrigues-Soares F, Naranjo MEG, Delgado Á, de Andrés F, *et al*. Interethnic variation of CYP2C19 alleles, 'predicted' phenotypes and 'measured' metabolic phenotypes across world populations. Pharmacogenomics J 2015; 16: 113–23.
- **5** Hsu HL, Woad KJ, Woodfield DG, Helsby NA. A high incidence of polymorphic CYP2C19 variants in archival blood samples from Papua New Guinea. Hum Genomics 2008; 3: 17–23.
- 6 Kaneko A, Kaneko O, Taleo G, Björkman A, Kobayakawa T. High frequencies of CYP2C19 mutations and poor metabolism of proguanil in Vanuatu. Lancet 1997; 349: 921–2.
- **7** Kaneko A, Lum JK, Yaviong J, Takahashi N, Ishizaki T, Bertilsson L, *et al.* High and variable frequencies of CYP2C19 mutations: medical consequences of poor drug metabolism in Vanuatu and other Pacific islands. Pharmacogenet Genomics 1999; 9: 581–90.
- **8** Kaneko A, Bergqvist Y, Taleo G, Kobayakawa T, Ishizaki T, Björkman A. Proguanil disposition and toxicity in malaria patients from Vanuatu with high frequencies of CYP2C19 mutations. Pharmacogenet Genomics 1999; 9: 317–26.

- 9 Masta A, Lum JK, Tsukahara T, Hwaihwanje I, Kaneko A, Paniu MM, et al. Analysis of Sepik populations of Papua New Guinea suggests an increase of CYP2C19 null allele frequencies during the colonization of Melanesia. Pharmacogenet Genomics 2003; 13: 697–700.
- **10** von Ahsen N, Tzvetkov M, Karunajeewa HA, Gomorrai S, Ura A, Brockmöller J, *et al.* CYP2D6 and CYP2C19 in Papua New Guinea: High frequency of previously uncharacterized CYP2D6 alleles and heterozygote excess. Int J Mol Epidemiol Genet 2010; 1: 310–19.
- **11** Griese EU, Ilett KF, Kitteringham NR, Eichelbaum M, Powell H, Spargo RM, *et al.* Allele and genotype frequencies of polymorphic cytochromes P4502D6, 2C19 and 2E1 in aborigines from western Australia. Pharmacogenet Genomics 2001: 11: 69–76.
- 12 Helsby N, Goldthorpe M, Gow P, de Zoysa J. Is the prevalence of CYP2C19 genetic variants different in Pacific people than in New Zealand Europeans? N Z Med J 2010; 123: 37–41.
- **13** Larsen PD, Johnston LR, Holley A, La Flamme AC, Smyth L, Chua EW, *et al.* Prevalence and significance of CYP2C19*2 and CYP2C19*17 alleles in a New Zealand acute coronary syndrome population. Intern Med J 2015; 45: 537–45.
- 14 Wanwimolruk S, Pratt EL, Denton JR, Chalcroft SCW, Barron PA, Broughton JR. Evidence for the polymorphic oxidation of debrisoquine and proguanil in a New Zealand Maori population. Pharmacogenet Genomics 1995; 5: 193–8.
- **15** Wanwimolruk S, Bhawan S, Coville PF, Chalcroft SCW. Genetic polymorphism of debrisoquine (CYP2D6) and proguanil (CYP2C19) in South Pacific Polynesian populations. Eur J Clin Pharmacol 1998; 54: 431–5.
- **16** Kaneko A, Bergqvist Y, Takechi M, Kalkoa M, Kaneko O, Kobayakawa T, *et al*. Intrinsic efficacy of proguanil against falciparum and vivax malaria independent of the metabolite cycloguanil. J Infect Dis 1999; 179: 974–9.
- 17 Zabalza M, Subirana I, Sala J, Lluis-Ganella C, Lucas G, Tomás M, *et al*. Meta-analyses of the association between cytochrome CYP2C19 loss- and gain-of-function polymorphisms and cardiovascular outcomes in patients with coronary artery disease treated with clopidogrel. Heart 2012; 98: 100–8.
- **18** Johnston LR, Larsen PD, La Flamme AC, Michel JM, Simmonds MB, Harding SA. Suboptimal response to clopidogrel and the effect of prasugrel in acute coronary syndromes. Int J Cardiol 2013; 167: 995–9.
- **19** Panattoni L, Brown PM, Te Ao B, Webster M, Gladding P. The cost effectiveness of genetic testing for CYP2C19 variants to guide thienopyridine treatment in patients with acute coronary syndromes. Pharmacoeconomics 2012; 30: 1067–84.
- 20 Bhopalwala AM, Hong RA, Khan ZR, Valentin MR, Badawi RA. Routine screening for CYP2C19 polymorphisms for patients being treated with clopidogrel is not recommended. Hawai'i J Med Public Health 2015; 74: 16–20.
- **21** Wu AH, White MJ, Oh S, Burchard E. The Hawaii clopidogrel lawsuit: the possible effect on clinical laboratory testing. Pers Med 2015; 12: 179–81.
- **22** Lea RA, Roberts RL, Green MR, Kennedy MA, Chambers GK. Allele frequency differences of cytochrome P450 polymorphisms in a sample of New Zealand Maori. N Z Med J 2008; 121: 33–7.

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- 23 Dunbar L, Miles W, Wheeler A, Sheridan J, Pulford J, Butler R. The CYP2D6 metaboliser status of patients prescribed risperidone for the treatment of psychosis. N Z Med J 2009; 122: 29-34.
- 24 LLerena A, Naranjo MEG, Rodrigues-Soares F, Penas-LLedó EM, Fariñas H, Tarazona-Santos E. Interethnic variability of CYP2D6
- alleles and of predicted and measured metabolic phenotypes across world populations. Expert Opin Drug Metab Toxicol 2014; 10: 1569-83.
- 25 Mehlotra RK, Ziats MN, Bockarie MJ, Zimmerman PA. Prevalence of CYP2B6 alleles in malaria-endemic populations of West Africa and Papua New Guinea. Eur J Clin Pharmacol 2006; 62: 267-75.